

CLINICAL APPROACH TO INFERTILE WOMEN AND THE VITRO FERTILIZATION (IVF): A MEDICINAL DISCIPLINE CHRONICLES

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ABSTRACT:

Before effective advice about treatment alternatives can be delivered, a complete yet focused history, physical exam, and evaluation of the infertile female patient is required. The initial appointment is not only for gathering information to guide proper testing, but it is also for developing a relationship with the patient, which supports the demanding nature of infertility therapy. This is critical when the patient undergoes stressful and sometimes time-consuming treatment, disappointment due to unmet expectations, and frequently financially taxing choices. After a thorough review or after other, less expensive reproductive treatments have failed; the choice to proceed to in vitro fertilization (IVF) may be made. In all cases, the patient may not be psychologically, emotionally, or financially prepared for IVF, emphasizing the need of building a favorable rapport as part of the infertile woman's or couple's pre-IVF evaluation. The main purpose of this Research Paper is to create an overview on the medical specialty chronicles a clinical approach to infertile women and in vitro fertilization (IVF).

Keywords: Hormonal imbalances, in vitro fertilization, infertile men and women, Laboratory layout.

INTRODUCTION:

Successful assisted reproduction requires close collaboration between doctors, scientists, nurses, and counselors, as well as the careful coordination of both a medical and a scientific approach to each couple who undergoes a treatment cycle. Only rigorous attention to detail at every stage of a patient's therapy can increase the likelihood of a healthy baby being born. Appropriate patient selection, ovarian stimulation, monitoring, and oocyte retrieval timing should supply viable gametes capable of creating healthy embryos to the in vitro fertilization (IVF) laboratory. The IVF laboratory is responsible for maintaining a stable, nontoxic, pathogen-free environment with optimal oocyte fertilization and embryo growth characteristics. The first half of this book delves into the complexities of the variables involved in ensuring successful fertilization and embryo development in animal systems, as well as the fascinating and exquisite molecular-level control systems. It goes without saying that human in vitro fertilization must entail systems of at least equal, if not greater, complexity, and it is critical for the clinical biologist to be aware that control mechanisms exist that are exquisitely sensitive to even seemingly minor changes in the environment of gametes and embryos, in particular, temperature, pH, and

any other factors that could potentially harm the gametes and embryos.

It goes without saying that human vitro fertilization must entail systems of at least equal, if not greater, complexity, and the clinical biologist must be aware that control mechanisms exist that are exquisitely sensitive to even seemingly minor changes in the environment of gametes and embryos, such as temperature, pH, and any other factors that may affect cells at the molecular level. Because there are so many variables, each step's basic science must be rigorously regulated while allowing for individual variance between patients and treatment cycles. Furthermore, in this day of fast expanding technology, the success of new techniques and technologies can only be measured against a criterion of efficient and effective performance. Furthermore, in today's world of quickly expanding technology, the success of new techniques and technologies can only be measured against a standard of efficient and repeatable established methods. The IVF laboratory thus has a responsibility and ability to respond not just to ensure that stringent cleanliness and sterile technique are followed throughout all procedures, but also to compile and maintain daily records with systematic data analysis and reporting.

Clinical Approach to the infertile man and woman:

Infertility is defined as the failure to conceive (be pregnant) after a year (or more) of unprotected sexual intercourse. Even though women's fertility declines with age, some doctors examine and treat women aged 35 and up after 6 months of unprotected intercourse. Women experiencing infertility should consult a reproductive endocrinologist, a doctor who specializes in infertility treatment. Women who have had two or more spontaneous miscarriages in the past may also benefit from

the services of reproductive endocrinologists. Pregnancy is the end consequence of a multi-step process. To become pregnant, follow these steps:

1. An egg must be released from one of a woman's ovaries external symbol.
2. Along the way, a man's sperm must unite with the egg (fertilize).
3. A fertilized egg must pass through a fallopian external symbol on its way to the uterus (womb).
4. The embryo must adhere to the uteri inside (implantation).

A difficulty with one or more of these processes can lead to infertility. Impaired fecundity is a type of infertility that affects women who have trouble getting pregnant or bringing a baby to term.

The purpose of taking a patient's history is to discover pregnancy contraindications, risk factors for infertility, and the necessity for referral to other services such as social work or anesthetic consultation. After a thorough examination, most individuals' causes of infertility will fall into one of many diagnostic categories that have been consistent over the last decade. The infertility expert is in a unique position to improve the patient's health prior to pregnancy and to provide preconception screening for diseases that can be passed down to the children, such as cystic fibrosis, spinal muscular atrophy, and other genetic disorders. Additionally, it is important information that after a year of trying, roughly 1 in 5 heterosexual women aged 15 to 49 years who have never given birth are unable to conceive (infertility). In addition, roughly one-fourth of the women in this category (26%) had trouble getting pregnant or bringing a pregnancy to term (impaired fecundity). Infertility and fecundity problems are less likely in women who have had one or more previous children. In this demographic, around 6% of married women aged 15 to 49 years are unable to

conceive after a year of trying, and 14% have trouble conceiving or carrying a pregnancy to term. The infertility expert is in a unique position to improve the patient's health prior to pregnancy and to provide preconception screening for diseases that can be passed down to the children, such as cystic fibrosis, spinal muscular atrophy, and other genetic disorders. While there are standard criteria for minimal preconception counseling and testing, there are fewer suggestions for management choices for specific reproductive disorders such as recurrent pregnancy loss or endometriosis. Both partners of a couple should be present for the initial consultation if at all possible, especially if treatment decisions are to be made.

On the other hand, Infertility in males can be caused by a variety of causes and is usually diagnosed through a sperm analysis. A specialist evaluates the amount of sperm (concentration), motility (movement), and morphology (shape) during a semen analysis. A little aberrant sperm analysis does not necessarily imply that a guy is infertile. A semen analysis, on the other hand, can assist evaluate if and how male variables contribute to infertility. The testicular or ejaculatory function is disrupted:

1. Varicocele, a condition in which a man's testicle's veins swell. Varicoceles can influence the amount and morphology of sperm, even if there are no symptoms.
2. Testicular trauma can impair sperm production and result in less sperm.
3. Heavy drinking, smoking, anabolic steroid use, and illicit drug use are all factors.
4. Chemotherapy, radiation, or surgery to remove one or both testicles as part of cancer treatment.
5. Testicular failure can be caused by medical illnesses such as diabetes, cystic fibrosis, certain autoimmune disorders, and infections.

Hormonal Imbalances:

The hypothalamus or pituitary glands aren't working properly. Hormones produced by the hypothalamus and pituitary glands in the brain keep testicular function normal. Low or no sperm production can be caused by excessive production of prolactin, a hormone produced by the pituitary gland (frequently due to the existence of a benign pituitary gland tumor), or other disorders that harm or impede the function of the brain or pituitary gland. Pituitary tumors, both benign and malignant (cancerous), congenital adrenal hyperplasia, too much estrogen, too much testosterone, Cushing's syndrome, and prolonged use of glucocorticoids are examples of these diseases. Genetic illnesses such as Klinefelter's syndrome, Y-chromosome microdeletion, myotonic dystrophy, and other, less prevalent genetic disorders can result in the production of no sperm or a small number of sperm.

The concept of In Vitro Fertilization (IVF):

IVF is a type of assisted reproduction in which a man's sperm and a woman's eggs are mixed outside of the body in a laboratory dish. One or more fertilized eggs (embryos) may be implanted in a woman's uterus, where they will implant and develop in the uterine lining. IVF medications and treatments rarely cause serious side effects. There are, however, some hazards associated with this treatment, as with any other medical procedure. The most common dangers are discussed in this document. In vitro fertilization (IVF) is the process of fertilizing a female egg outside the body in a laboratory with male sperm, often in a cell culture dish. The fertilized egg can then be put in the female's uterus, where it will attempt to attach and develop. The eggs, sperm, or embryos may be frozen, preserved, and stored for later use in some instances.

Setting up An IVF laboratory and basics: Equipment and facilities for setting up a

laboratory With rational and logical planning, an IVF laboratory should be designed to create a distraction- and accident-free atmosphere in which undivided attention can be comfortably and safely given to each manipulation. With work stations that are practical and easy to clean. The tissue culture area should allow for the highest standards of sterile technique, with all floors, surfaces, and components easy to clean on a regular basis, as well as minimizing the potential for introducing infection or contamination from any source. The location should ideally be identified as a restricted access area, with facilities for changing into clean operating theater attire and shoes before to admission.

Air quality in the environment Cohen et al. examined the importance of ambient air and the potential repercussions of chemical air contamination (1997). Unlike other creatures and species, oocytes and embryos in vitro lack such protection, and their active and passive absorption processes are generally indiscriminate. IVF laboratories located in polluted locations or near industrial manufacturing facilities may be subjected to substantial chemical air contamination, which might result in low pregnancy and live birth rates. CO₂ is supplied in gas bottles, which may be polluted with organic compounds or metallic impurities. Incubators get their ambient air straight from the laboratory room; CO₂ is supplied in gas bottles, which may be contaminated with organic compounds or metallic contaminants. Many IVF laboratories utilize pressurized rooms with high efficiency particulate air (HEPA) filtration, which meets pharmaceutical clean room standards; nevertheless, HEPA filtration cannot successfully retain gaseous low molecular weight organic and inorganic compounds. The four most common contaminants in the air are:

1. VOCs created by industry, a range of cleaning techniques, and car and heating

exhausts in densely populated urban and suburban areas. VOCs can also be produced by instruments like microscopes, television monitors, and furniture (due to manufacturing procedures).

2. N₂O, SO₂, and CO is examples of small inorganic compounds.
3. Building-material-derived substances, such as aldehydes from flooring adhesives, substituted benzenes, phenol, and n-decane released from vinyl floor tiles - flooring adhesives have been shown to be particularly active in halting embryo development! Many paints include compounds that are particularly hazardous in the IVF lab; therefore freshly painted surfaces are frequently a hazard.
4. Pesticides or aerosols employing butane or iso-butane as a propellant may release other polluting substances. Heavy metals can be found in liquids like floor waxes, which have a significant impact on embryo implantation potential.

Cohen and colleagues investigated chemical air contamination in all parts of their IVF lab, uncovering dynamic interacting processes between air-handling systems, spaces, instruments, disposable materials, and other laboratory-specific things. Anesthetic gases, refrigerants, cleaning agents, hydrocarbons, and aromatic chemicals were discovered, with some accumulating in incubators especially. They propose that there may be an interaction between water-soluble and lipid-soluble solid phases, such as those found in incubators: while some pollutants may be absorbed by culture media, this can be mitigated by providing a larger sink, such as a humidification pan. Mineral oil has the potential to behave as a sink for other substances. Due to the accumulation of VOCs from surrounding rooms or certain laboratory items, such as sterile Petri dishes, unfiltered outdoor air may be cleaner than HEPA filtered

laboratory air or air received from incubators. Manufacturers of compressed air and incubators are unconcerned about the unique clean air requirements of IVF: compressed gas supply standards are based on criteria that are inappropriate for cultured and unprotected cells. Allowing the emission of gases from new laboratory products is critical. Testing a new incubator revealed quantities of VOC > 100-fold greater than those obtained from testing used incubators from the same manufacturer. Devoted that existing air-conditioning methods appear incapable of preventing pollution from the environment within the laboratory, main concern should be given to the architecture of culture spaces and neighboring regions. Active filtration units with activated carbon filters and oxidizing material have now been created specifically for IVF laboratories to avoid the problem of potential risks in ambient air and culture systems.

Laboratory Space Layout:

To reduce the risk of accidents, careful consideration should be paid to the physical maneuvers involved, ensuring ease and safety of mobility between locations. Bench height, adjustable seats, microscope eye height, and effective use of space and surfaces all contribute to a distraction-free working environment. Within each working space, the placement of storage rooms and laboratory equipment such as incubators and centrifuges should be logically arranged for efficiency and safety; the use of transportable laboratory components offers flexibility to meet changing requirements. The following is a list of the essential equipment needed for normal IVF:

1. Light, inverted, and dissecting microscopes.
2. Incubator with temperature and CO₂ levels that are precisely controlled.
3. For sperm preparation, use a centrifuge.
4. Culture manipulations on warmed stages or surfaces.

5. Freezer/refrigerator

6. For drying and sterilization, use a dry heat oven.

For teaching, assessment, and records, a video camera system is also recommended (patients receive enormous satisfaction and psychological support from observing their oocytes and embryos on a video screen). When selecting these costly pieces of equipment, be sure they are not only simple to operate and maintain, but also that servicing and repairs are available swiftly and efficiently. Cleaning, maintenance, and servicing schedules must be developed for each piece of equipment, and checklist records must be kept for daily, weekly, monthly, and annual cleaning and maintenance plans for all items utilized, as well as checks for restocking and supply expiration dates.

There are many other reasons behind the intensification man's risk of infertility, such as: Age, although advanced age is a better predictor of female infertility, couples with a male partner who is 40 years old or older are more likely to have trouble conceiving. Obesity is being overweight. Smoking, Excessive use of alcohol and other drugs (opioids, marijuana). Testosterone exposure. When a doctor provides testosterone injections, implants, or topical gel for low testosterone, or when a guy consumes testosterone or related medicines illegally to increase muscle mass, this can happen. Radiation exposure. Frequent exposure of the testes to high temperatures, such as that experienced by men who are confined to a wheelchair or who use a sauna or hot tub on a regular basis. Flutamide, cyproterone, bicalutamide, spironolactone, ketoconazole, or cimetidines are some of the drugs that can cause this.

When IVF Is the Next Treatment Option: Preparing an infertile woman for IVF necessitates a thorough discussion and consent

procedure that must be documented and include the following:

1. The steps of an IVF cycle, including how the ovaries will be stimulated, pharmaceutical side effects, follicular maturation management, monitoring, and the period of gonadotropin stimulation expected.
2. IVF alternatives
3. Surgical and anesthetic risks associated with oocyte retrieval
4. Risk of ovarian torsion and ovarian hyper stimulation syndrome
5. Current information on the risk of cancer as a result of treatment, if any.
6. The possibility of genetic disease, congenital malformations, or other diseases in the infant as a result of IVF.
7. Pregnancy and failure rates expected as a result of age-related decreased fecundity and increased aneuploidy.
8. Ectopic pregnancy, early delivery, low birth weight, and placentation anomalies are some of the pregnancy issues encountered following IVF.
9. Insemination or intracytoplasmic sperm injection on a regular basis (ICSI).
10. The number of embryos to be transferred and the length of the culture.
11. When alternative treatments, such as assisted hatching and preimplantation screening, should be investigated.
12. The possibility of numerous pregnancies.
13. There is no assurance of success.
14. When IVF fails or when using a donor oocyte for IVF is necessary.

Evaluation of Oocyte Quality:

A level of more than 25 mIU/mL (about 12 mIU/mL using current assays) has been linked to a poor likelihood of pregnancy. Recent research has found that small elevations in the pregnancy rate predict a more modest reduction in the pregnancy rate in women under the age of 40, however an

elevated level has much more relevance in older women. Follicular maturation can happen quickly, especially in older women, and the FSH level can start to drop by day 3. As a result, the level of estradiol (E2) should be examined as well. It's unknown what effect an elevated day 3 E2 level (more than 70–80 pg/mL) has in the presence of a normal FSH concentration. Because the ovarian response has been demonstrated to fluctuate inversely with the FSH level, FSH levels can be utilized to estimate the appropriate level of stimulation. We usually use a low responder strategy for women with an FSH level more than 10 mIU/mL. The normal ranges of FSH tests vary greatly. A level of 12 mIU/mL corresponded to 25 mIU/mL with the assay used in the aforementioned report when we converted to the Immulite system (Siemens Healthcare Global, Erlangen, Germany). If there isn't a direct comparison, the College of American Pathologists survey booklet can be used, which offers mean levels for all laboratories employing each kit and standard sera (College of American Pathologists, Northfield, Illinois).

FSH levels differ from one cycle to the next. A regularly increased FSH level predicts a worse prognosis than a single elevated level with normal levels elsewhere. When IVF is performed during a cycle with a lower FSH level, the quality of ovarian stimulation does not increase. Although there is agreement that women with a single elevated FSH level have a higher cancellation risk, studies disagree on the extent to which the pregnancy outcome is reduced. FSH levels are generally identical on days 2, 3, and 4 of the cycle, yet in someone with fast follicular development, day 2 may be the ideal day to disclose an increased FSH level due to a prior elevated serum estradiol. The first day of complete flow should be counted as day 1 for women with premenstrual spotting.

Antral Follicle Count (AFC):

The antral follicles respond to stimulation, as measured by the antral follicle count (AFC). These can be reliably counted with a high-quality trans-vaginal ultrasound scan. In healthy women, AFC declines with age. A typical response to stimulation is expected in women with 5–10 follicles each side. When there are more than 10 follicles on each side (polycystic ovary [PCO]-like), a lower dose of stimulation should be utilized than would be used normally based on weight and FSH level. A low AFC (less than 5 or 6 total) indicates a poor prognosis and should induce the implementation of a low responder strategy and increased stimulation levels. AFC is favorably correlated with the number of retrieved oocytes and negatively with day 3 FSH and gonadotropin ampoules, with fewer than 10 total follicles indicating a higher risk of cancellation. AFC was found to be a greater predictor of ovarian response than FSH in multivariate analysis. Because the success rate of IVF is so low in women over 40 who grow fewer than three follicles with stimulation, a low AFC can be used with other data (age, day 3 FSH, and infertility duration) to propose egg donation as a better choice.

Antimullerian Hormone (AMH):

AMH is a greater predictor of ovarian response and live birth than AFC, and it can be evaluated at any point during the menstrual cycle. When corrected for age, AMH, like AFC, provides some limited prediction for successful pregnancy. AMH levels that are very low or undetectable have been linked to a low but adequate ovarian response and pregnancies in some women, and should not be used to rule out stimulation. The AMH level is increasingly being utilized to determine the optimal level of stimulation; one study found that a level of > 3.36 ng/mL accurately predicted OHSS with over 90% sensitivity.

Related Construction to ovarian reserve testing:

The CCCT has been used to identify patients with a poor prognosis and ovarian reserve who have a normal day 3 FSH level. AFC and AMH have overtaken this test for predicting ovarian response and IVF success. With stimulation, women with PCOS develop more follicles. Because more oocytes are extracted, the fertilization rate is reduced. The pregnancy rate is comparable to that of other IVF patients. Metformin enhanced implantation and lowered the miscarriage incidence by 50% in a meta-analysis of ten randomized studies. Although the odds ratio for birth was 1.69, it did not reveal a statistically significant trend. The risk of ovarian hyper stimulation syndrome (OHSS) was reduced by more than 70% (odds ratio 0.27, CI 0.16–0.46). Metformin has been found to minimize the follicular and estradiol response to stimulation, increase the number of mature oocytes and embryo quality, and raise the pregnancy rate in clomiphene-resistant women with PCOS. Insulin resistance is more common in clomiphene-resistant women; therefore, this clinical category and insulin resistance may be very strong indicators for this supplementary treatment. Women with PCO who coasted on metformin had lower peak estradiol levels and fewer days of coasting. Metformin is a crucial aid in lowering OHSS in these women since insulin is one of the key variables that stimulates luteinized granulosa cell production of vascular endothelial growth factor, and metformin lowers ovarian response and circulating insulin levels. As it is necessary to know that as a woman's age has become the leading cause of infertility, ovarian reserve testing (ORT), or reproductive potential as a function of the number and quality of surviving oocytes, is becoming more relevant. Many middle-aged women seek the help of reproductive specialists in the hopes of overcoming the

negative consequences of aging using assisted reproduction procedures (ART). However, the reality is much different, and physicians must advise patients about their actual chances of conception before commencing ART. ART has become a big issue in this environment. At weeks 15–18 of pregnancy, the ovaries contain 6–7 million oocytes surrounded by a layer of granulosa cells, forming the primordial follicle pool. Only 1–2 million primordial nongrowing follicles (PNGFs) are left at birth.

At menarche, the ovaries contain 300,000–400,000 PNGFs, which decrease to less than 1000 at menopause over the reproductive years. Only 12 percent of the original ovarian reserve is kept through the age of 30, and only 3% by the age of 40, according to studies. Regression analyses of two relevant databases have recently attempted to establish the age of menopause in women based on the number of resting PNGFs. Ultrasonographic features that identify antral follicles or detect granulosa cell products of growing follicles in serum are used to assess ovarian reserve, reflecting a population of follicles with a specific degree of development and reactivity to gonadotropins. Only 12 percent of the original ovarian reserve is kept through the age of 30, and only 3% by the age of 40, according to studies. Regression analyses of two relevant databases have recently attempted to establish the age of menopause in women based on the number of resting PNGFs. Ultrasonographic features that identify antral follicles or detect granulosa cell products of growing follicles in serum are used to assess ovarian reserve, reflecting a population of follicles with a specific degree of development and reactivity to gonadotropins. Because there is another true ovarian reserve of PNGFs that is not recognized by these markers, this is not the true ovarian reserve.

A drop in follicle number is accompanied by a drop in oocyte quality. This is thought to be related to a rise in meiotic nondisjunction,

which leads to an increase in aneuploidy in the early embryo as the embryo ages. Human aneuploidy is a major factor in the aging of the human reproductive system. Differences in quality between germ cells at the moment of formation, accumulated oocyte damage throughout the course of a woman's life, or age-related changes in the quality of the surrounding granulosa cells that influence the oocyte could all be underlying processes. The best indicator of quality is age. In fact, monthly fecundity begins to decline around the age of 30, and natural fertility loss (measured by age at last child in conditions of unrestricted reproduction) begins around the age of 41, with a range of 23 to 51 years of age. However, women's reproductive aging differs significantly, with some remaining fertile into their fifth decade of life and others losing fertility in their mid-thirties. Women over 35 who have not conceived after 6 months of trying to conceive are candidates for ART due to the normal fall in fertility with age. Furthermore, based on their clinical history, a group of patients can be identified as having a higher risk of reduced ovarian reserve (DOR). ART markers that are suitable are highly repeatable, widely available, noninvasive, and cost-effective. They should also be able to predict the birth rate. Efforts have been made to find the optimum approach to test it, but neither single nor combined markers are sufficient at the moment.

In Vitro Maturation Clinical Indications:

1. Cohort of Patients: The major goal of IVF treatment in most clinics is to help women with polycystic ovaries (PCO) avoid ovarian hyperstimulation syndrome (OHSS). This is a significant medical condition in which the lady not only suffers from acute discomfort as a result of the rapid development of ascites, but she also runs the risk of a thrombotic event. IVF is now used to treat a variety of

reproductive concerns in patients, including fertility preservation, hormone-sensitive malignancies, FSH resistance, and cost-conscious individuals.

2. Polycystic Ovaries (PCO) and Polycystic Ovarian Syndrome (PCOS) are two types of polycystic ovaries.

OHSS is a serious clinical consequence of gonadotropin stimulation in women with PCO/PCOS, resulting in patient discomfort in the moderate stages and considerable morbidity in the more severe forms. The only therapy option that entirely removes the risk of OHSS is IVM. The antral follicle count is a primary driver of IVM therapy success rates, and patients with PCOS often have a high antral follicle count and hence respond better to treatment. PCOS patients often have more oocytes harvested from IVM cycles than non-PCOS patients, resulting in a higher pregnancy rate.

3. Women with regular menstrual cycles without PCO or PCOS: are more controversial to include in IVM treatment regimens, and because the antral follicle count is the determining criterion for IVM treatment, patients with fewer than five antral follicles should not be considered. With this in mind, IVM has been shown to be a viable therapeutic option for non-PCOS patients with adequate antral follicle levels. The literature comparing patient cohorts is confined to those evaluating stimulation techniques, with patient selection criteria appearing as a by-product of success, and more study in this area is needed.

4. Fertility Preservation: Oncofertility is a new area of assisted reproductive technology (ART) in which patients must retain their gametes and embryos prior to treatment for a variety of cancers that put their future fertility at risk. IVM is a feasible oncofertility treatment option for patients with estrogen-sensitive malignancies, as well as a mechanism for maturing ex vivo oocytes acquired during

oophorectomy or ovarian tissue cryopreservation.

5. Estrogen Sensitive Cancer/Avoidance of OHSS: Prior to chemo/radiotherapy, the most frequent malignancy in women seeking fertility preservation is breast cancer. IVM is a safer alternative to controlled ovarian hyperstimulation in patients with estrogen-sensitive cancers since it can be conducted without the use of exogenous gonadotropins, reducing the danger of increased circulating estrogen levels, and it may be their only therapy choice. Additionally, IVM may be a good alternative for patients whose cancer is not estrogen-sensitive because it fully removes the danger of OHSS, allowing them to start chemo/radiotherapy right away without having to wait for life-saving cancer therapies.

6. The Time of Oophorectomy or Ovarian Tissue Cryopreservation: In patients with Hodgkin's lymphoma, breast cancer, and rectal cancer, IVM has been effectively performed during oophorectomy and laparoscopic ovarian wedge excision for ovarian tissue cryopreservation. Immature oocytes were extracted from visible follicles in the lab prior to ovarian cortex tissue cryopreservation, with all patients storing at least one mature oocyte and ovarian tissue cryopreservation successfully. This procedure gives the patient two possibilities for future reproductive support. IVM was effectively conducted with oocytes harvested ex vivo following laparoscopic oophorectomy in a recent case report. 22 GV stage oocytes were retrieved from both ovaries, with 15 mature oocytes vitrified after 24 hours in maturation culture and four more vitrified after another 24 hours.

Although it is unknown whether oocyte survival, fertilization, or development will be successful, a large number of cryopreserved oocytes will give the patient the best chance of future fertility. There has been a live birth after in vitro maturation of oocytes collected after

oophorectomy. However, oocyte vitrification in standard IVF treatment significantly improves oocyte survival, fertilization, and embryo quality, as well as clinical and ongoing pregnancy rates, when compared to slow freezing techniques, and hopefully this will translate to oocytes collected from IVM cycles. Oocyte cryopreservation, rather than embryo cryopreservation, is required in situations where the patient is not currently in a relationship and time is of the essence, and IVM is frequently the only treatment capable of attaining this goal.

7. Conversion from IVF: For patients who begin to recruit too many follicles, conversion from IVF to IVM may be conducted to avoid cycle termination or OHSS. This strategy was found to be effective in patients seeking fertility preservation, as these patients' cycles would have been cancelled or their cancer treatment would have been delayed if they had not used it. Instead, they kept a large number of mature oocytes and embryos for future fertility. This procedure can be applied to other patients as a way to avoid OHSS if the patient is properly advised and gives their consent to change their treatment type.

8. Avoidance of OHSS: in addition to oncofertility therapies, IVM can be utilized to avert OHSS in some circumstances. In rare cases, such as when a patient lives in a rural or remote place, a treatment like IVM allows the patient to fly home immediately after treatment, reducing the risk of OHSS or a thrombotic event, which is not normally recommended after standard IVF treatment. This could also be effective in circumstances when infertility is not a concern, such as for a young egg donor with a high antral follicle count, and a minimalist approach to stimulation is preferred to minimize excessive side effects from ovarian stimulation. While there is no indication that IVM was used in

these situations, it is available in clinics that do IVM on a regular basis.

9. Prothrombotic Risk: It is widely known that a thrombotic event following IVF treatment is a severe cause of morbidity, due to the high estradiol released as part of an IVF cycle activating the clotting cascade, which may be aggravated by the hemoconcentration associated with OHSS. By maintaining a low circulating estrogen level and removing the risk of OHSS, IVM has the potential to lower the risk of prothrombotic diseases. There have yet to be any cases of IVM being employed when a prothrombotic disease was the determining factor. However, if the antral follicle count is sufficient to indicate IVM as a viable treatment option, there is no reason to believe this protocol will fail.

10. IVM for FSH Resistant Ovaries: In a patient with FSH-resistant ovaries, a pregnancy and live birth were obtained. On a patient with early onset ovarian failure who was sent to the clinic for donor oocytes, the researchers used their normal IVM approach. With a sufficient number of antral follicles, IVM was a viable option, with maturation, fertilization, and embryo development rates equivalent to those seen in other IVM patients at the clinic. The success of this case demonstrates the value of having IVM as a treatment option in ART clinics where oocyte donation is the only other viable alternative for the patient.

Important missive to previous discussion: The amount of gap junction communication between the cumulus cells and the oocyte decreases significantly when the cumulus-oocyte complex is removed from the follicle, which must be regulated in order to prevent spontaneous maturation and promote synchronization between nuclear and cytoplasmic maturation. For usage in IVM, various culture media have been developed. Sage (Cooper Surgical, USA) and Medicult (Origio, Denmark), the two most extensively

used commercially available IVM base media, were shown to be equally effective. When compared to Sage media, specifically designed media for blastocyst culture have proven to be equally effective in terms of maturation, fertilization, and blastocyst development. In recent years, research has focused on factors that influence embryo development and a variety of additives, including an exogenous protein source, growth factors, steroids, energy sources, hormones, oocyte secreted factors, activators, and inhibitors, have been tested with varying degrees of success. Hormonal additions are now a standard component of all human IVM culture media. There is a rapid and large fall in cAMP concentrations after immature oocytes are removed from follicles, which is hypothesized to cause asynchrony between oocyte nuclear and cytoplasmic maturation.

Growth factors are less common in oocyte maturation culture media and should be utilized with caution due to the unknown effects on epigenetic variation. Some researchers have proposed that adding insulin-like growth factor (IGF-I) to the culture conditions encourages cumulus cell enlargement and nuclear maturation through promoting granulosa cell division. Similarly, epidermal growth factor (EGF) has been demonstrated to help with GV breakdown and polar body extrusion maturation rates. When cultured with cumulus intact oocytes, EGF has also been demonstrated to help in fertilization. In vitro maturation rates of oocytes were also boosted by EGF family members' recombinant human Areg and Ereg. In the IVM culture media, a protein supply is required. Human follicular fluid (HFF), inactivated autologous patient serum (human serum), or fetal bovine serum are the three main sources of protein in humans (FBS). HFF has been utilized at concentrations ranging from 30% to 70%. Patient serum is used at a concentration of

10% or 20%, with inactivated FBS at a concentration of 10% or 20%. Human serum supplies a number of nutrients and factors that are important in the maturation process that are not always present in synthetic serums. However, typical IVF embryo culture has a variety of disadvantages for developing embryos, as well as the potential of introducing unknown contaminants into the cultured embryos when utilizing serum or HFF preparations. It's unclear whether serum's detrimental effects in embryo culture also apply to oocyte maturation culture. Depending on the strategy used, the oocytes are only exposed to the maternal serum for the first 24–48 hours before being transferred to commercial embryo culture media and inseminated. Furthermore, the culture environment for in vitro oocyte maturation is designed to imitate the in vivo intra-follicular milieu, and follicular fluid and serum contain many of the same components.

In traditional IVF, steps are being taken to imitate the in vivo environment by using low oxygen tension, which is similar to what the embryo experiences in vivo. While many laboratories throughout the world continue to employ ambient oxygen levels, the ART field as a whole agrees that low oxygen tension enhances clinical pregnancy and live birth rates. As a result, most centers use low oxygen tension in their IVM culture as well. However, no studies comparing oxygen concentration during in vitro maturation on human oocytes have been published. The hardening of the zona pellucida was thought to necessitate intracytoplasmic sperm injection (ICSI) in oocytes acquired after IVM culture due to the longer maturation culture schedule. Although fertilization rates were lower in the IVF inseminated group compared to the ICSI inseminated group, embryo cleavage and implantation rates were greater for PCO and PCOS patients, according to the first study to

examine insemination procedures following IVM treatment. There was no difference in fertilization, usable blastocyst development, total blastocyst development, or implantation rates between the two insemination strategies in a sibling oocyte research comparing fertilization techniques in IVM oocytes. Even while using sister oocytes to compare procedures reduces confounding factors that can affect results, there is a potential error in the timing of maturation/fertilization checks performed between ICSI and IVF. Large-scale randomized control trials are clearly needed for IVM to be deemed a validated therapy option for ART clinics. Furthermore, more research into the long-term outcomes of children born using this procedure is required. IVM is more time-consuming logistically than regular IVF, due to the extra time required for in vitro culture, but a more consistent approach to protocols around the world could lead to a wider adoption of the treatment. Additional culture media additives, such as oocyte secreted factors GDF9 and BMP-15, as well as cAMP modulators (IBMX), have showed promise in animal experiments, while IBMX's efficacy in terms of embryo chromosomal aneuploidy has been proven in human volunteers.

The most typical culture period for human clinical IVM embryos was two to three days after insemination until recently. However, blastocyst culture is becoming more common in IVF, and the determining factor for culture times of IVF generated embryos should reflect the clinics performing the technique's standard culture methods. Time lapse incubation systems are now being used for IVF as a result of recent advancements in technology for embryo culture systems. Analyses of embryo development using time lapse monitoring of embryo development found an increase in some abnormal phenotypic events in PCOSIVM embryos.

Furthermore, IVM embryos were shown to have a higher rate of early arrest; however, IVM treatment had no effect on the morphokinetic development of embryos suitable for frozen transfer. More large-scale research exploring its impact on human IVF success rates could lead to the production of more successful, commercially available culture medium, bringing IVM closer to the success rates seen with standard IVF. With success rates for IVM procedures improving and data demonstrating positive outcomes in children conceived after this treatment, routine use of IVM in clinical practice may not be a long way off. Laboratory practices will benefit from the creation of commercial culture media to increase success rates and the standardization of protocols. More thorough investigation into the effectiveness of IVM treatment is still needed.

Concluding Remark:

In vitro maturation (IVM) is an alternative to in vitro fertilization (IVF) in which the patient receives little or no gonadotropin stimulation and the harvested oocytes are matured in the laboratory. After oocytes obtained from unstimulated follicles underwent spontaneous maturation, it was first documented in an animal model and then duplicated in a human model. After oocyte maturation following ovarian biopsy, the first live birth was recorded in 1991, followed by trans-vaginal methods in 1994. The major goal of IVM treatment is to prevent the negative effects and higher expenditures associated with follicle stimulating hormone medication (FSH). IVM, on the other hand, has the ability to overcome other causes of infertility, such as gamete donation, FSH resistance, avoiding the consequences of a high estradiol level, and fertility preservation. More than 3000 infants are said to have been born as a result of this approach globally, and while certain clinics use

it on a regular basis, it is still regarded a research technique. This research paper aims to look into patient cohorts, treatment and laboratory protocols, success rates, and birth outcomes reported from clinics doing IVM all around the world, as well as areas that need more research and future trends for IVM. The paucity of randomized control studies and long-term data on the outcomes of infants born using this method, in particular. IVM, like regular IVF, falls perfectly into this category as a financially profitable and patient-friendly treatment option as many lines of science advance toward a more sustainable, least intrusive approach.

Abbreviation:

Co₂: Carbon di oxide

HEPA: High-Efficiency Particulate Air

VOC's: Vaso-Occlusive Crisis

N₂o: Nitrous Oxide

So₂: Sulfur dioxide

CO: Carbon mono oxide

ICSI: Intracytoplasmic sperm injection

FSH: Follicle Stimulating Hormone

AFC: Antral Follicle Count

PCOS: Polycystic Ovary Syndrome

PCO: Polycystic Ovary

OHSS: Ovarian Hyper stimulation Syndrome

CCCT: Clomiphene Citrate Challenge Test

ART: Antiretroviral therapy

IVM: In-vitro maturation

GV: Germinal Vesicle

EGF: Epidermal growth factor

HFF: Hypertensive Heart Failure

IBMX: 3-isobutyl-1-methylxanthine

REFERENCES:

- 1) Keck, C., Fischer, R., Baukloh, V., & Alper, M. (2005). Staff management in the in vitro fertilization laboratory. *Fertility and sterility*, 84(6), 1786-1788.
- 2) Brison, D. R., Hooper, M., Critchlow, J. D., Hunter, H. R., Arnesen, R., Lloyd, A., &

Horne, G. (2004). Reducing risk in the IVF laboratory: implementation of a double witnessing system. *Clinical Risk*, 10(5), 176-180.

- 3) Matson, P. L. (1998). Internal quality control and external quality assurance in the IVF laboratory. *Human Reproduction*, 13(suppl_4), 156-165.
- 4) Montag, M. H., & Morbeck, D. E. (Eds.). (2017). *Principles of IVF laboratory practice: Optimizing performance and outcomes*. Cambridge University Press.
- 5) Gianaroli, L., Plachot, M., van Kooij, R., Al-Hasani, S., Dawson, K., DeVos, A., ... & Embryology, C. O. T. S. I. G. O. (2000). ESHRE guidelines for good practice in IVF laboratories. *Human Reproduction*, 15(10), 2241-2246.
- 6) Simopoulou, M., Sfakianoudis, K., Maziotis, E., Grigoriadis, S., Giannelou, P., Rapani, A., ... & Koutsilieris, M. (2019). The impact of autoantibodies on IVF treatment and outcome: a systematic review. *International journal of molecular sciences*, 20(4), 892.
- 7) Wu, L. H., Humm, K. C., Dodge, L. E., Sakkas, D., Hacker, M. R., & Penzias, A. S. (2012). IVF outcomes are paradoxically poorer under age 25. *Fertility and Sterility*, 98(3), S264.
- 8) Margalioth, E. J., Ben-Chetrit, A., Gal, M., & Eldar-Geva, T. (2006). Investigation and treatment of repeated implantation failure following IVF-ET. *Human reproduction*, 21(12), 3036-3043.
- 9) Forte, M., Faustini, F., Maggiulli, R., Scarica, C., Romano, S., Ottolini, C., ... & Rienzi, L. (2016). Electronic witness system in IVF—patients perspective. *Journal of assisted reproduction and genetics*, 33(9), 1215-1222.
- 10) Ni, L., Sadiq, S., Mao, Y., Cui, Y., Wang, W., & Liu, J. (2013). Influence of various tubal surgeries to serum antimullerian hormone level and outcome of the subsequent IVF-ET

- treatment. *Gynecological Endocrinology*, 29(4), 345-349.
- 11) Siristatidis, C., Stavros, S., Drakeley, A., Bettocchi, S., Pouliakis, A., Drakakis, P., ... & Vlahos, N. (2021). Omics and Artificial Intelligence to Improve In Vitro Fertilization (IVF) Success: A Proposed Protocol. *Diagnostics*, 11(5), 743.
- 12) Jubaer, S. M. O. F., & Ahmed, J. Deficiency in Evidence Law Concerning Technological and Expert Support. *JournalNX*, 7(05), 1-10.
- 13) Van den Bergh, M., Hohl, M. K., De Geyter, C., Stalberg, A. M., & Limoni, C. (2005). Ten years of Swiss National IVF Register FIVNAT-CH. Are we making progress?. *Reproductive biomedicine online*, 11(5), 632-640.
- 14) Van den Bergh, M., Hohl, M. K., De Geyter, C., Stalberg, A. M., & Limoni, C. (2005). Ten years of Swiss National IVF Register FIVNAT-CH. Are we making progress?. *Reproductive biomedicine online*, 11(5), 632-640.
- 15) Group, C. C. (2020). 'There is only one thing that is truly important in an IVF laboratory: everything' Cairo Consensus Guidelines on IVF Culture Conditions. *Reproductive BioMedicine Online*, 40(1), 33-60.
- 16) Schlegel, P. N., Berkeley, A. S., Goldstein, M., Cohen, J., Alikani, M., Adler, A., ... & Rosenwaks, Z. (1994). Epididymal micropuncture with in vitro fertilization and oocyte micromanipulation for the treatment of unreconstructable obstructive azoospermia. *Fertility and sterility*, 61(5), 895-901.
- 17) Schlegel, P. N., Berkeley, A. S., Goldstein, M., Cohen, J., Alikani, M., Adler, A., ... & Rosenwaks, Z. (1994). Epididymal micropuncture with in vitro fertilization and oocyte micromanipulation for the treatment of unreconstructable obstructive azoospermia. *Fertility and sterility*, 61(5), 895-901.
- 18) Suikkari, A. M. (2008). In-vitro maturation: its role in fertility treatment. *Current Opinion in Obstetrics and Gynecology*, 20(3), 242-248.
- 19) Gibreel, A., El-Adawi, N., Elgindy, E., Al-Inany, H., Allakany, N., & Tournaye, H. (2015). Endometrial scratching for women with previous IVF failure undergoing IVF treatment. *Gynecological Endocrinology*, 31(4), 313-316.
- 20) Guo, T., Qin, Y., Gao, X., Chen, H., Li, G., Ma, J., & Chen, Z. J. (2012). The role of male chromosomal polymorphism played in spermatogenesis and the outcome of IVF/ICSI-ET treatment. *International journal of andrology*, 35(6), 802-809.
- 21) Jubaer, S. M. O. F., & Hassan, M. N. (2021). THE ARTIFICIAL INTELLIGENCE FOR THE COMMON LEARNERS: A COMPARATIVE LEARNING APPROACH. *Web of Scientist: International Scientific Research Journal*, 2(05), 333-3525.
- 22) Sneed, M. L., Uhler, M. L., Grotjan, H. E., Rapisarda, J. J., Lederer, K. J., & Beltsos, A. N. (2008). Body mass index: impact on IVF success appears age-related. *Human reproduction*, 23(8), 1835-1839.
- 23) Jubaer, S., & Hoque, L. (2021). The Concept of Education: A Western Rationalist Approach. *International Journal of Educational Advancement*, 4, 138-150.
- 24) Sadeghi, M. R. (2018). The 40th anniversary of IVF: has ART's success reached its peak?. *Journal of Reproduction & Infertility*, 19(2), 67.
- 25) Jubaer, S. M. O. F., Joly, J. A., Devi, A. D., & Shovon, A. I. (2021). WORK PARTICIPATION OF FEMALES AND EMERGING LABOR CIRCUMSTANCES IN BANGLADESH. *Journal of Marriage and the Family*, 949, 959.
- 26) Balaban, B., Barut, T., & Urman, B. (2012). Assessment of oocyte quality. In *Practical*

- Manual of in Vitro Fertilization* (pp. 105-119). Springer, New York, NY.
- 27) Hsu, A., Arny, M., Knee, A. B., Bell, C., Cook, E., Novak, A. L., & Grow, D. R. (2011). Antral follicle count in clinical practice: analyzing clinical relevance. *Fertility and sterility*, 95(2), 474-479.
- 28) Hendriks, D. J., Mol, B. W. J., Bancsi, L. F., Te Velde, E. R., & Broekmans, F. J. (2005). Antral follicle count in the prediction of poor ovarian response and pregnancy after in vitro fertilization: a meta-analysis and comparison with basal follicle-stimulating hormone level. *Fertility and sterility*, 83(2), 291-301.
- 29) Galtier-Dereure, F., De Bouard, V., Picot, M. C., Vergnes, C., Humeau, C., Bringer, J., & Hedon, B. (1996). Endocrinology: Ovarian reserve test with the gonadotrophin-releasing hormone agonist buserelein: correlation with in-vitro fertilization outcome. *Human reproduction*, 11(7), 1393-1398.
- 30) Bancsi, L. F., Broekmans, F. J., Eijkemans, M. J., de Jong, F. H., Habbema, J. D. F., & te Velde, E. R. (2002). Predictors of poor ovarian response in in vitro fertilization: a prospective study comparing basal markers of ovarian reserve. *Fertility and sterility*, 77(2), 328-336.
- 31) La Marca, A., Papaleo, E., Grisendi, V., Argento, C., Giuliani, S., & Volpe, A. (2012). Development of a nomogram based on markers of ovarian reserve for the individualisation of the follicle-stimulating hormone starting dose in in vitro fertilisation cycles. *BJOG: An International Journal of Obstetrics & Gynaecology*, 119(10), 1171-1179.
- 32) Broer, S. L., Broekmans, F. J., Laven, J. S., & Fauser, B. C. (2014). Anti-Müllerian hormone: ovarian reserve testing and its potential clinical implications. *Human reproduction update*, 20(5), 688-701.
- 33) Kyweluk, M. (2019). *Ovarian Reserve Testing in the United States* (Doctoral dissertation, Northwestern University).
- 34) Depmann, M., Broer, S. L., Van Der Schouw, Y. T., Tehrani, F. R., Eijkemans, M. J., Mol, B. W., & Broekmans, F. J. (2016). Can we predict age at natural menopause using ovarian reserve tests or mother's age at menopause? A systematic literature review. *Menopause*, 23(2), 224-232.
- 35) Depmann, M., Broer, S. L., Van Der Schouw, Y. T., Tehrani, F. R., Eijkemans, M. J., Mol, B. W., & Broekmans, F. J. (2016). Can we predict age at natural menopause using ovarian reserve tests or mother's age at menopause? A systematic literature review. *Menopause*, 23(2), 224-232.
- 36) Walls, M. L., & Hart, R. J. (2018). In vitro maturation. *Best practice & research Clinical obstetrics & gynaecology*, 53, 60-72.
- 37) Hatırnaz, Ş., Ata, B., Hatırnaz, E. S., Dahan, M. H., Tannus, S., Tan, J., & Tan, S. L. (2018). Oocyte in vitro maturation: A systematic review. *Turkish Journal of Obstetrics and Gynecology*, 15(2), 112.
- 38) Hatırnaz, Ş., Ata, B., Hatırnaz, E. S., Dahan, M. H., Tannus, S., Tan, J., & Tan, S. L. (2018). Oocyte in vitro maturation: A systematic review. *Turkish Journal of Obstetrics and Gynecology*, 15(2), 112.
- 39) Hoshino, Y. (2018). Updating the markers for oocyte quality evaluation: intracellular temperature as a new index. *Reproductive medicine and biology*, 17(4), 434-441.
- 40) Lasienė, K., Vitkus, A., Valanėiūtė, A., & Lasys, V. (2009). Morphological criteria of oocyte quality. *Medicina*, 45(7), 509.
- 41) Bianchi, L., Gagliardi, A., Campanella, G., Landi, C., Capaldo, A., Carleo, A., ... & Bini, L. (2013). A methodological and functional proteomic approach of human follicular fluid en route for oocyte quality evaluation. *Journal of proteomics*, 90, 61-76. Chicago

- 42) Boni, R. (2018). Origins and effects of oocyte quality in cattle. *Animal Reproduction (AR)*, 9(3), 333-340.
- 43) Uchikura, K., Nagano, M., & Hishinuma, M. (2010). Evaluation of follicular development and oocyte quality in pre-pubertal cats. *Reproduction in Domestic Animals*, 45(6), e405-e411.
- 44) Sciorio, R., Miranian, D., & Smith, G. D. (2022). Non-invasive oocyte quality assessment. *Biology of Reproduction*, 106(2), 274-290.
- 45) Jubaer, S. M. O. F., & Ahmed, J. Deficiency in Evidence Law Concerning Technological and Expert Support. *JournalNX*, 7(05), 1-10.
- 46) Jubaer, S. M. O. F., & Hassan, M. N. (2021). The political ideology and philosophy of Bangabandhu Sheikh Mujibur Rahman in the context of founding a nation. *World Bulletin of Social Sciences*, 2, 24-35.
- 47) Murakawa, H., Aono, N., Tanaka, T., Kikuchi, H., Yoshida, H., Yoshida, H., ... & Abe, H. (2009). Morphological Evaluation and Measurement of the Respiration Activity of Cumulus-oocyte complexes to Assess oocyte quality. *Journal of Mammalian Ova Research*, 26(1), 32-41.
- 48) Lehmann, P., Vélez, M. P., Saumet, J., Lapensée, L., Jamal, W., Bissonnette, F., ... & Kadoch, I. J. (2014). Anti-Müllerian hormone (AMH): a reliable biomarker of oocyte quality in IVF. *Journal of assisted reproduction and genetics*, 31(4), 493-498.
- 49) Schlegel, P. N., Berkeley, A. S., Goldstein, M., Cohen, J., Alikani, M., Adler, A., ... & Rosenwaks, Z. (1994). Epididymal micropuncture with in vitro fertilization and oocyte micromanipulation for the treatment of unreconstructable obstructive azoospermia. *Fertility and sterility*, 61(5), 895-901.